

Overview

We present a semi-automatic workflow for characterizing antibody primary structure utilizing protein contigs assembled from *de novo* peptide sequences.

Introduction

De novo peptide sequencing is a practical technique for characterizing uncharted proteins. Among all proteins, antibodies are the most common subject to be sequenced. In general, it is required to achieve complete sequence coverage with multiple enzymatic digestion and produce evidence for every residue, specifically for the hypervariable CDR regions. While automated *de novo* peptide sequencing is routinely conducted, it is still puzzling and tedious to infer the original protein sequence from those short peptides.

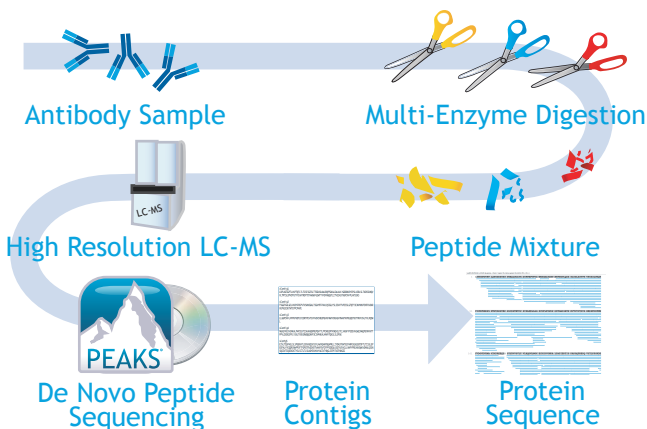


Figure 1. Workflow of Monoclonal Antibody Characterization

In the world of genomics, DNA sequencing is achieved by assembling short reads using software. In principle, one can do the same in proteomics by assembling the short *de novo* peptides into “protein contigs”, and it will significantly reduce the workload in manual analysis.

Method

1. Assembling of protein contigs

A program is developed using a greedy algorithm for assembling protein contigs. It first detects the overlapping *de novo* peptides and iteratively extends the contigs using the consensus residues at both direction.

In the extending process, contigs are merged if their sequences collide with each other. Protein contigs are extended until no more residues can be confidently added.

N-terminus of a protein contig

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HYAPVTYLQVE.....
  GHYAPVTYLE
  TVPYAHVGDGHYAPVTYLQVE
  ATSDHYAPVTFGAE
  EAPGTVINANVDGHYAPVTYLQVE
  TPGTF LADQHGHYAPVTYLKAD
  GHYAPVTFKE
  TTGDHYAPVTFKE
  GHYAPVTYLGAVE
  MLLSMGVAYATHYAPVTFQE
  TTGDHYAPVTYLE
    
```

Supporting *de novo* peptides

Figure 2. Extending a Contig at N-Terminus

2. Workflow for antibody sequencing

1. Assemble an antibody template

- Blast protein contigs in NCBI nr database
- Select a protein hit corresponding to the constant region
- Select the closest protein hit corresponding to the variable region.

2. Draft a sequence using the contigs and template

- Map the contigs to the antibody template
- Solve disagreements between contigs and the template
 - ♦ Correct typical *de novo* sequencing errors (I/L, AG/Q, PK/KP, variable PTMs)
 - ♦ In principle, trust contigs in variable region and the template in constant region
- Solve the residues not covered by contigs, if any, using *de novo* peptides

3. Refine the sequence using PEAKS' SPIDER homology search

- Perform SPIDER homology search on the draft sequence
- Examine insertion/deletion/mutation reported by SPIDER
- Examine residues with low peptide coverage
- Examine residues at the protein n-terminus
- Compare the sequence mass with protein intact mass, if available
- Make corrections and repeat step 3 until the sequences converge

Result

1. Experiment

The antibody, I5154 (human IgG1/kappa), was purchased from Sigma-Aldrich to evaluate the proposed workflow. The sample was reduced with DTT, alkylated with iodoacetamide. Glycans were removed and heavy/light chains were separated. Each chain was digested with six enzymes AspN, chymotrypsin, GluC, LysC, pepsin and trypsin. MS/MS spectra was acquired using LTQ-Orbitrap at high resolution with HCD fragmentation.

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The mass spec data for the heavy and light chains were separately analyzed. *De novo* peptide sequencing was performed using PEAKS. Protein contigs were assembled from *de novo* peptides. The majority of assembled contigs had a length of 60-120 residues. Antibody sequences were manually constructed using the described workflow. For both the heavy and light chains, sequencing results were fully covered by MS/MS spectra. 98% of amino acid residues were supported by fragment ions in at least 5 MS/MS spectra.



Figure 3. Coverage of a Light Chain Sequencing Result

2. Blind trials

The proposed workflow was also tested in a series of blind trials. Our collaborators provide antibody mass spec data to us for in-house analysis, while not disclosing the true sequences. The sequencing results were correct at a minimal of 96% of residues, meanwhile all the wrong residues were previously identified with low confidence during the sequencing analysis.

Conclusion

Protein sequencing can be efficiently conducted by utilizing protein contigs assembled from *de novo* peptides.

The described semi-automated workflow provides a reliable solution for antibody sequencing with MS/MS of multiple enzymatic digestion.

The proposed workflow has been routinely used for data analysis in the **CHAMPS** - Antibody Sequencing Service, provided by Bioinformatics Solution Inc. (<http://www.bioinfor.com/peaks/products/champs.html>)