SILAC-based Quantitative Post Translational Modification (PTM) Profile Analysis with PEAKS

Wen Zhang, Ziaur Rahman, Yifang Chen, Lei Xin, and Baozhen Shan
Bioinformatics Solutions Inc., Waterloo, Canada

Aims

To overview SILAC (Stable Isotope Labeling with Amino Acids in Cell Culture)-based PTM analysis in PEAKS Q.

Summary:

PEAKS supports complex SILAC data analysis and quantitative PTM profiling of individual protein and all identified proteins. A published dataset of SILAC-based comparative quantification of phosphoproteome was used to demonstrate SILAC data analysis and PTM Profile function in PEAKS Studio.

Benefits

- Accurate and sensitive detection and association of SILAC feature pairs
- Transfer IDs between associated SILAC pairs and different MS runs
- Flexible experimental design and statistical tools to facilitate data analysis

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Introduction

➢ SILAC and Super-SILAC

SILAC (stable isotope labeling by amino acids in cell culture) has become one of the most popular labeling techniques for mass spectrometry (MS)-based quantitative proteomics [1]. In this metabolic labeling strategy, differential isotope labeled samples (proteins/peptides) are combined early in the experimental procedure and analyzed together by LC -MS/MS. Therefore, the variation introduced from sample processing is minimized. Since stable-isotope labeled peptides have almost the same physicochemical properties as their natural counterparts, the same peptides with differential labeling co-elute from the liquid chromatography (LC) column and their amounts can be accurately quantified relative to each other.

An extension of this labeling strategy is named super-SILAC [2, 3], where labeled samples can be produced separately and spiked into each of the experimental samples that are not amenable to metabolic labeling, e.g. human tissues. This pool of spiked-in heavy proteins is used as an internal standard for quantification.

PEAKS Q supports analyses of SILAC and super-SILAC types of data.

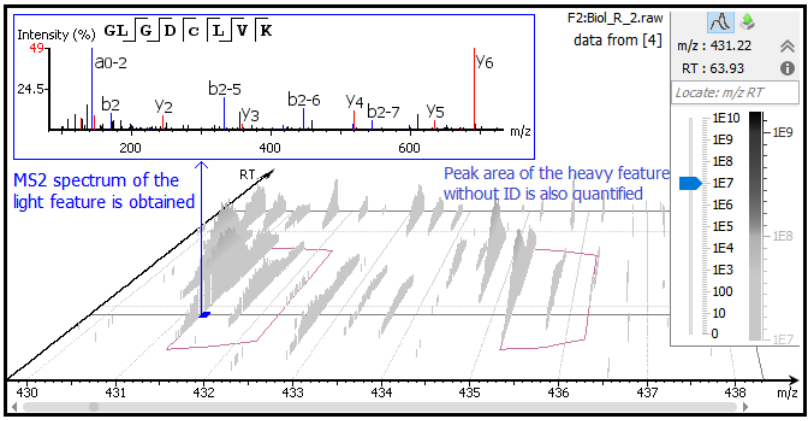
➢ PTM

Protein post-translational modifications (PTMs) play critical roles in diverse biological processes. Identification and characterization of PTMs are essential for a comprehensive understanding of cellular biology and human diseases. Recently, MS-based isotope labeling strategies such as SILAC, TMT and iTRAQ have gained popularity in quantitative proteomics, which have been applied to identify and quantify PTMs successfully. A new PTM Profiling function has been implemented in PEAKS Studio software for PTM analysis and direct visualization of results.

SILAC Quantification Algorithms in PEAKS Q

➢ ID transfer between associated SILAC pairs

PEAKS Q detects and associates 2 or 3-plex SILAC feature pairs that have the same charge, similar MS1 peak area correlation over retention time, expected mass shifts caused by labeling and fall within certain mass errors. If an identification is obtained from one of the



3 feature vectors

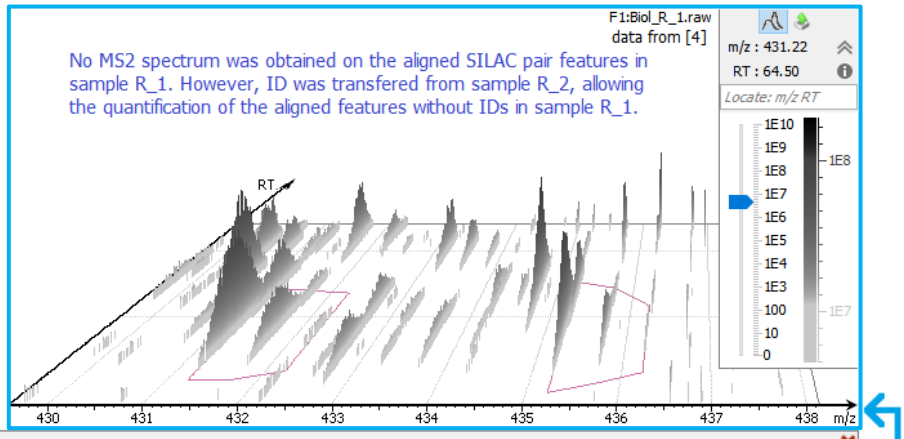
	Sample	Fraction	-10lgP	Quality Score	Light_Area	Heavy_Area	Light_Ratio	Heavy_Ratio	Id Count	m/z	z	ppm	RT	AScore
1	R_1	F 1	20.18	60.60	2.47E8	2.42E8	1.00	0.98	0	431.2292	2	1.5	63.93	
2	R_2	F 2	20.18	59.12	2.75E8	2.62E8	1.00	0.95	1	431.2292	2	1.5	63.93	
3	R_2_1	F 3	25.06	63.84	1.89E8	1.92E8	1.00	1.02	2	431.2288	2	0.5	64.30	

Peptide	-10lgP	Quality Score	Heavy : Ratio Profile	Group 1 : Light_Ratio	Group 1 : Heavy_Ratio	#Vector	Accession
GLGDC(+57.02)LVK(*)	25.06	61.19		1.00	0.98	3	P12236|ADT3_HUMAN:P05141|ADT2_HUMAN

labeled states, then the whole SILAC pair feature can be quantified and used for peptide and protein ratio calculations. For example, the light form of peptide GLGDCLVK was fragmented and identified in sample R_2 [4]. Although no MS2 spectrum was obtained from the K8 labeled heavy counterpart, the SILAC pair could still be quantified in PEAKS Q as highlighted in the feature vectors table. “Id Count” indicates the number of MS2 identified for each SILAC pair.

➢ ID transfer between different MS runs after alignment

Retention times of different LC-MS runs are first aligned, then the MS/MS and ID can be matched from another run by aligning features within tight mass ranges and retention times, allowing quantification of SILAC pairs without any ID.



3 feature vectors

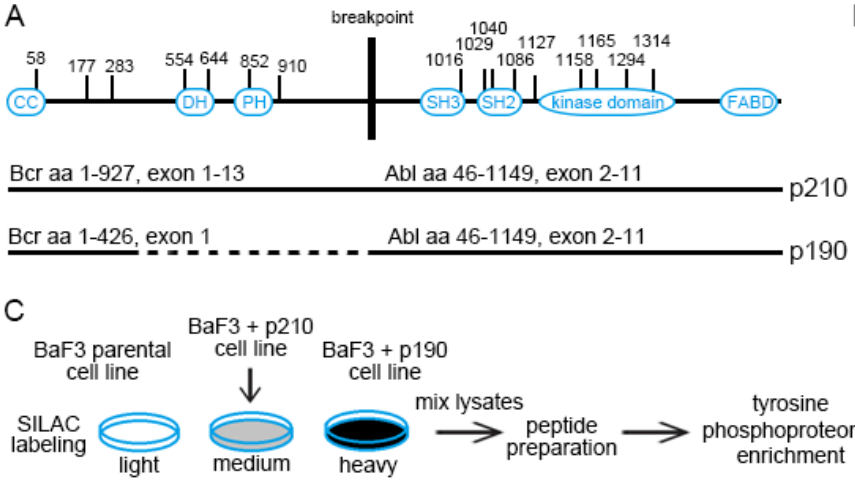
	Sample	Fraction	-10lgP	Quality Score	Control_Area	Treated_Area	Control_Ratio	Treated_Ratio	Id Count	m/z	z	ppm	RT	AScore
1	R_1	F 1	20.18	60.60	2.47E8	2.42E8	1.00	0.98	0	431.2292	2	1.5	63.93	
2	R_2	F 2	20.18	59.12	2.75E8	2.62E8	1.00	0.95	1	431.2292	2	1.5	63.93	
3	R_2_1	F 3	25.06	63.84	1.89E8	1.92E8	1.00	1.02	2	431.2288	2	0.5	64.30	

Case Study

A published dataset [5] was used as an example to demonstrate the capability of SILAC-based data analysis (PTM profiling) in PEAKS Studio.

➢ Study Aims and Background

The two major isoforms of the oncogenic Bcr–Abl tyrosine kinase, p210 and p190, were suggested to contribute to different types of leukemia. One of the main goals of the study was to identify differential signaling networks of Bcr-Abl p210 and p190 kinases in leukemia cells by using quantitative proteomics approaches [5]. Bcr–Abl is a fusion protein and a constitutively active tyrosine kinase. p190 is ~25% shorter than p210 due to a lack of a DH–PH domain unit; otherwise p210 and p190 have an identical sequence and domain organization (Figure 1A).



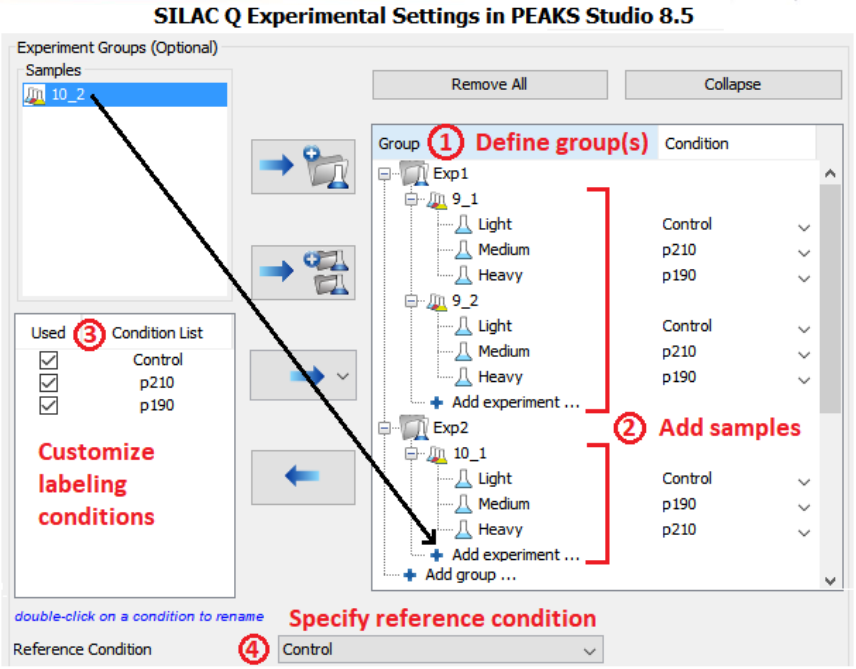
B

Experimental Design	
Exp 1: Sample 9 × 2 replicates	
Light	Control
Medium	p210
Heavy	p190
Exp 2: Sample 10 × 2 replicates	
Light	Control
Medium	p190
Heavy	p210

Figure 1. Bcr-Abl protein domain organization (A) and overview of the phosphoproteome experiments (B, C).

➢ Experimental Design

Murine BaF3 cells that express Abl endogenously were retrovirally transduced with the human Bcr–Abl p210 and p190 cDNAs. Parental (untransduced) BaF3 cells were used as a control and grown in SILAC light media (Figure 1C). Cells that expressed human p210 and p190 were labeled with SILAC medium and heavy media and swapped between 2 experiments (defined as Exp 1 and 2 in Figure 1B). Phosphotyrosine peptides were enriched and analyzed by high-resolution LC-MS/MS.



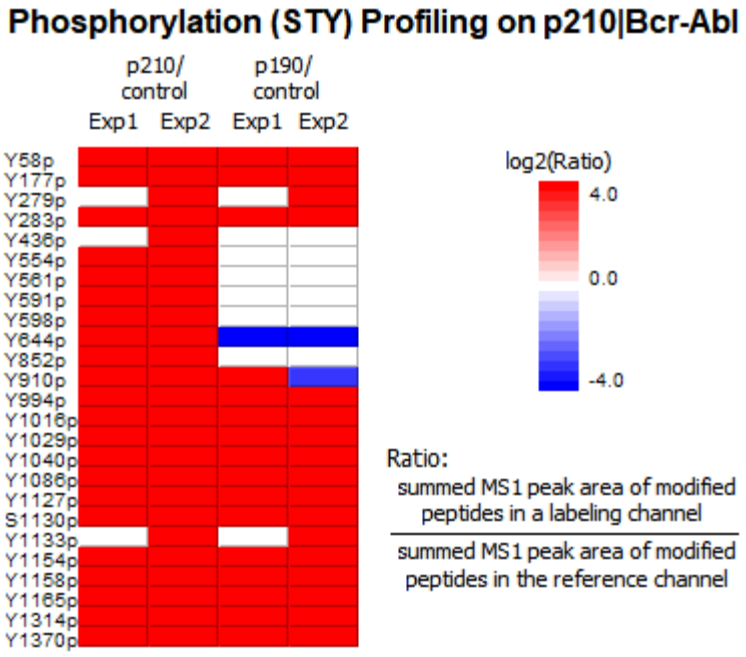
➢ Data analysis

MS data was analyzed in PEAKS Studio 8.5 using the built-in SILAC-3plex (R6, K4|R10, K8) method in PEAKS Q for quantification.

Results

➢ The Phosphorylation Profile of p210/p190 with PEAKS PTM Profile

The phosphorylation profile of Bcr-Abl proteins were analyzed with the PTM Profile function in PEAKS. 25 phosphorylation sites of Bcr-Abl were quantified (Ascore [6] > 20). As expected p210 and p190 samples showed higher phosphorylation signals compared to controls. For each confident PTM site, MS1 peak area of modified peptides are summed for each labeling channel and ratios relative to the reference channel are calculated automatically and displayed in the heatmap, enabling easy comparative quantitation of PTM abundances between samples and across groups. Furthermore, PTM quantification results can be normalized by protein expression levels automatically.



Conclusions

PEAKS Studio supports complex SILAC data analysis and quantitative PTM profiling analysis.

References

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